

# NGS sample specifications - WGS

Library Type	Sample Type	Input	Volume	Amount	
WGS	TruSeq DNA Nano	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 20 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 15 \mu\text{l}$ TE-Buffer (max. 50 $\mu\text{l}$ )	at least 300 ng in 15 $\mu\text{l}$
	Illumina DNA prep	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 1 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 10\mu\text{l}$ TE-Buffer (max. 55 $\mu\text{l}$ )	at least 5 ng in 10 $\mu\text{l}$
	Illumina DNA PCR-Free	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 5 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 10\mu\text{l}$ TE-Buffer (max. 55 $\mu\text{l}$ )	At least 50 ng in 10 $\mu\text{l}$
	Sequel II SMRTBell HIFI	gDNA (double-stranded – <i>high molecular weight</i> )	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 100 \text{ ng}/\mu\text{l} \leq 500 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 20 \mu\text{l}$ TE-Buffer (max. 40 $\mu\text{l}$ )	at least 1.5 $\mu\text{g}/\text{Gb}$ haploid genome in 20 $\mu\text{l}$
	Sequel SMRTBell microbial multiplexed genomes	gDNA (double-stranded – <i>high molecular weight</i> )	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 100 \text{ ng}/\mu\text{l} \leq 500 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 20 \mu\text{l}$ TE-Buffer (max. 40 $\mu\text{l}$ )	at least 1 $\mu\text{g}$ in 20 $\mu\text{l}$

# NGS sample specifications – targeted sequencing

	Library Type	Sample Type	Input	Volume	Amount
Targeted Sequencing	Illumina DNA Prep with Enrichment IDT xGen Exome	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 10 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 15 \mu\text{l}$ TE-Buffer (max. $50 \mu\text{l}$ )	at least 150 ng in $15 \mu\text{l}$
	Custom panels (contact first)	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 10 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 15 \mu\text{l}$ TE-Buffer (max. $50 \mu\text{l}$ )	at least 150 ng in $15 \mu\text{l}$
	HLA typing	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 30 \text{ ng}/\mu\text{l} \leq 500 \text{ ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 40 \mu\text{l}$ TE-Buffer (max. $100 \mu\text{l}$ )	at least 1 200 ng in $40 \mu\text{l}$

# NGS sample specifications – methylome sequencing

	Library Type	Sample Type	Input	Volume	Amount
Methylome sequencing	RRBS	gDNA (double-stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 20 \text{ ng}/\mu\text{l} \leq 200</math></u> <u><math>\text{ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 10\mu\text{l}$ TE-Buffer (max. $25\mu\text{l}$ )	at least 200 ng in $10 \mu\text{l}$
	NEB Em-Seq	gDNA (double stranded)	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 1 \text{ ng}/\mu\text{l} \leq 200\text{ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 15\mu\text{l}$ TE-Buffer (max. $55\mu\text{l}$ )	at least 15 ng in $15 \mu\text{l}$
	Sequel SMRTBell	gDNA (double-stranded – <i>high molecular weight</i> )	$OD_{260/280} \geq 1,8$ <u>concentration <math>\geq 150 \text{ ng}/\mu\text{l} \leq 500</math></u> <u><math>\text{ng}/\mu\text{l}</math></u> (based on Qubit)	in $\geq 20 \mu\text{l}$ TE-Buffer (max. $40 \mu\text{l}$ )	at least $1.5 \mu\text{g}/\text{Gb}$ haploid genome in $20 \mu\text{l}$

# NGS sample specifications – self made libraries and microbiome sequencing

	Library Type	Sample Type	Input	Volume	Amount
selfmade libraries	Ready to sequence libraries (P5/P7 adapter ligated)	any	<b>concentration <math>\geq 10\text{ng}/\mu\text{l} \leq 200\text{ng}/\mu\text{l}</math> or <math>&gt; 2 \text{ nM}</math></b> (based on Qubit)	in $\geq 15 \mu\text{l}$ TE-Buffer	-
	Amplicon PacBio SMRTBell (up to 8 kb)	Amplicon with <a href="#">universal overhang</a>	PCR product of first round of amplification described in this <a href="#">pdf</a> <b>concentration <math>\geq 1 \text{ ng}/\mu\text{l} \leq 10\text{ng}/\mu\text{l}</math></b> (based on Qubit AFTER clean up)	in $\geq 15 \mu\text{l}$ Tris-Buffer	-
Microbiome/Amplicon	Shotgun Metagenome Sequencing	gDNA (double stranded)	$\text{OD}_{260/280} \geq 1,8$ <b>concentration <math>\geq 1 \text{ ng}/\mu\text{l} \leq 200 \text{ ng}/\mu\text{l}</math></b> (based on Qubit)	in $\geq 10 \mu\text{l}$ TE-Buffer (max. 55 $\mu\text{l}$ )	at least 5 ng in 10 $\mu\text{l}$
	Bacterial 16S Amplicon V1/V2	}	Please contact Corinna Bang (C.bang@ikmb.uni-kiel.de)		
	Bacterial 16S Amplicon V3/V4				
	Archaeal 16S Amplicon				
Fungal ITS					

# NGS sample specifications – transcriptome sequencing

	Library Type	Sample Type	Input	Volume	Amount
RNA libraries	TruSeq stranded RNA	total RNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 40 ng/μl ≤ 200ng/μl</b> (based on Qubit)	in ≥ 15 μl TE-Buffer (max. 30 μl)	at least 600 ng in 15μl
		mRNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 40 ng/μl ≤ 200ng/μl</b> (based on Qubit)	in ≥ 15 μl TE-Buffer (max. 50 μl)	at least 600 ng in 15μl
	Illumina stranded RNA	Total RNA with RiboZero plus (H/M/R + bacteria)	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 10 ng/μl ≤ 200ng/μl</b> (based on Qubit)	in ≥ 15 μl TE-Buffer (max. 30 μl)	at least 150 ng in 15μl
		mRNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 2 ng/μl ≤ 200ng/μl</b> (based on Qubit)	in ≥ 15 μl TE-Buffer (max. 30 μl)	at least 30 ng in 15μl
	NextFlex smallRNA	total RNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 0.5 ng/μl</b> (based on Qubit)	in 15 μl TE-Buffer	at least 7.5 ng in 15 μl
		micro RNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) <b>Amount ≥ micro RNA fraction of ≥ 1,2 μg total RNA</b> (based on Qubit)	in 15 μl TE-Buffer	-
	PacBio IsoSeq	total RNA	DNA free pure (OD <sub>260/280</sub> ≥ 1,8) RIN (Agilent Bioanalyzer 2100) ≥ 8 <b>concentration ≥ 150 ng/μl ≤ 400ng/μl</b> (based on Qubit)	in ≥ 10 μl Tris Buffer or nuclease free water	at least 800 ng in 10 μl

# NGS sample specifications – single cell sequencing

	Library Type	Sample Type	Input	Volume	Amount
	Chromium single-cell RNAseq (contact us before)	Cell suspension	<b>&gt; 500 Cells/sample</b> Viability at least 95 %	in $\geq 10 \mu\text{l}$ buffer (max. 40 $\mu\text{l}$ )	-